

## On Gap Junctions

### What is a Gap Junction?

- Cluster of gap junction channels
- Linking structure between neighbouring cells
- Provides direct passage of molecules and ions

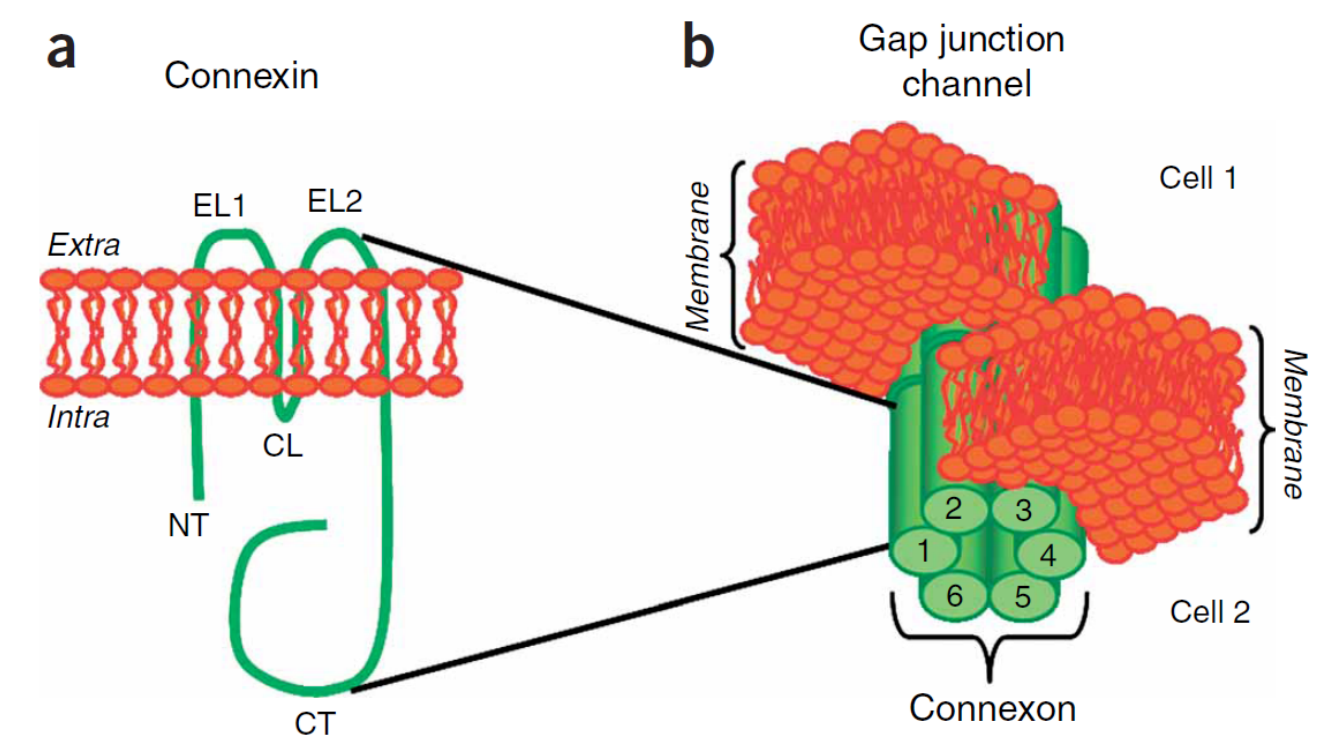


Figure : Schematic diagrams of a standard connexin molecule and gap junction channel. (Del Corso et. al., 2006)

### What are they made of?

- Proteins connexins.
- 6 connexins = 1 connexon (hemichannel)
- 2 connexons = 1 gap junction channel
- Cardiac cell proteins: Cx43, Cx45 and Cx40.

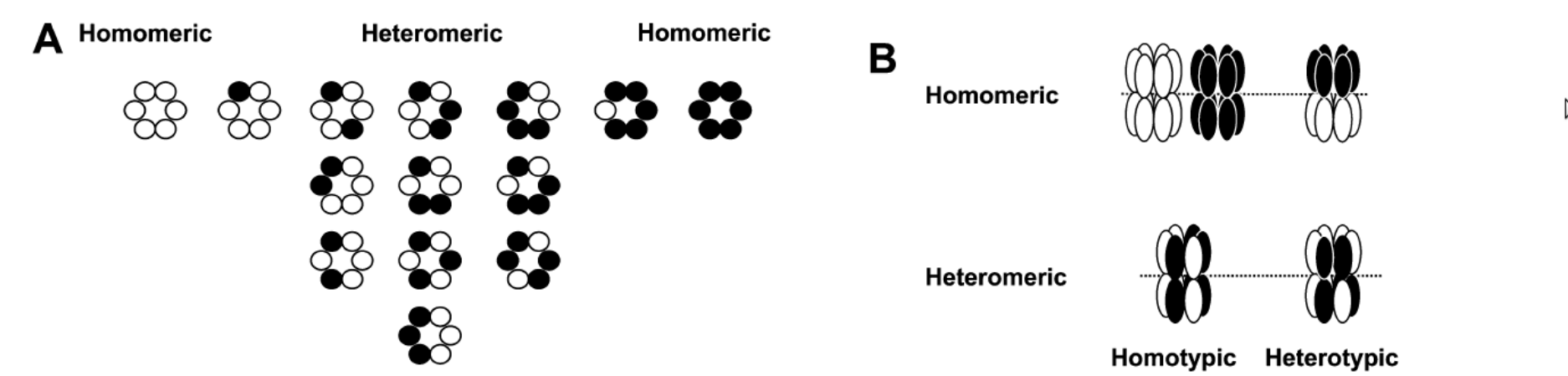


Figure : Predicted configurations of connexons and gap junction channels for two different connexins. (Desplantez, 2004)

### Where are they located in cardiac cells?

- Mostly on the longitudinal ends where they compose the intercalated disks
- The behaviour of transversal GJ channels is not well understood

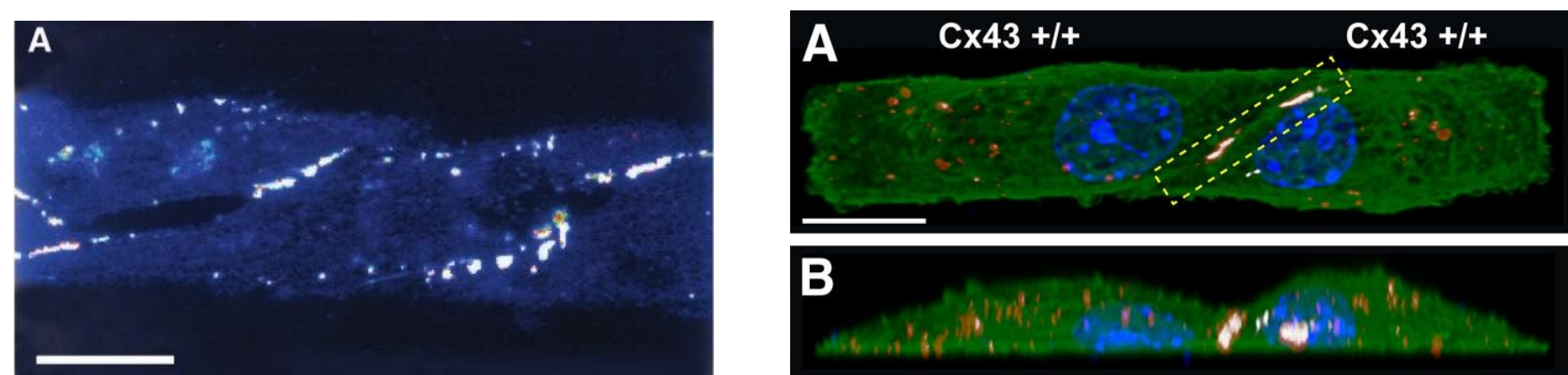
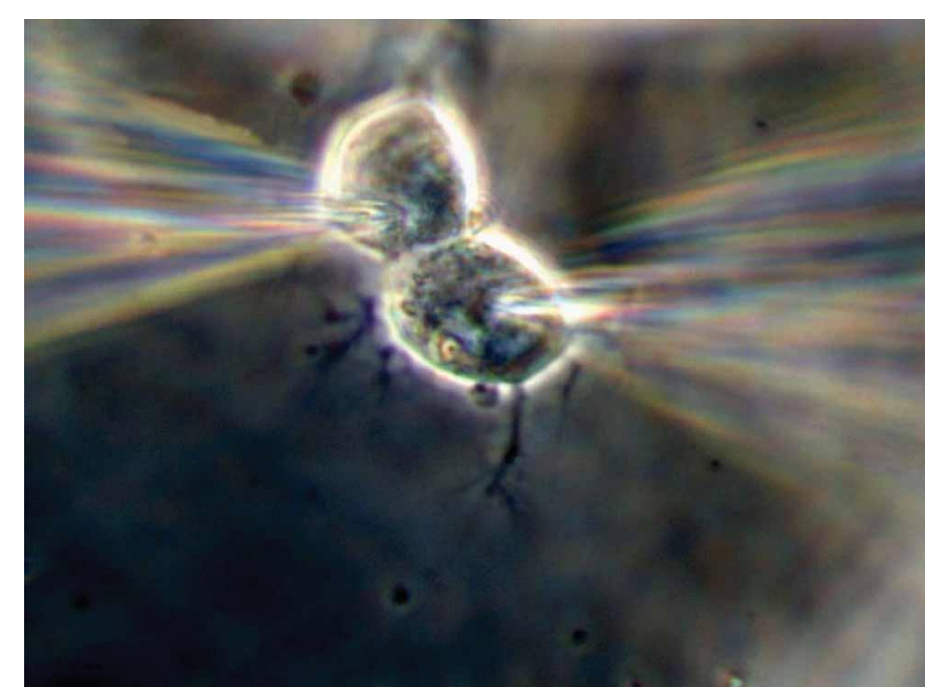


Figure : Immunohistochemical analysis of Cx43. Left: Bar = 10 μm. (Beauchamp, 2004) Right: top view(A), lateral view(B). Bar = 5 μm. (Beauchamp, 2012)

### Electrical behaviour?

- The dual voltage patch clamp
- Non-linear behaviour - gating of channels
- Dependent on connexin arrangement.



## Non linear GJ model - 0D

- GJ current:  $I_j = G_j(t, V_j)V_j$ , where  $V_j$  transjunctional voltage.

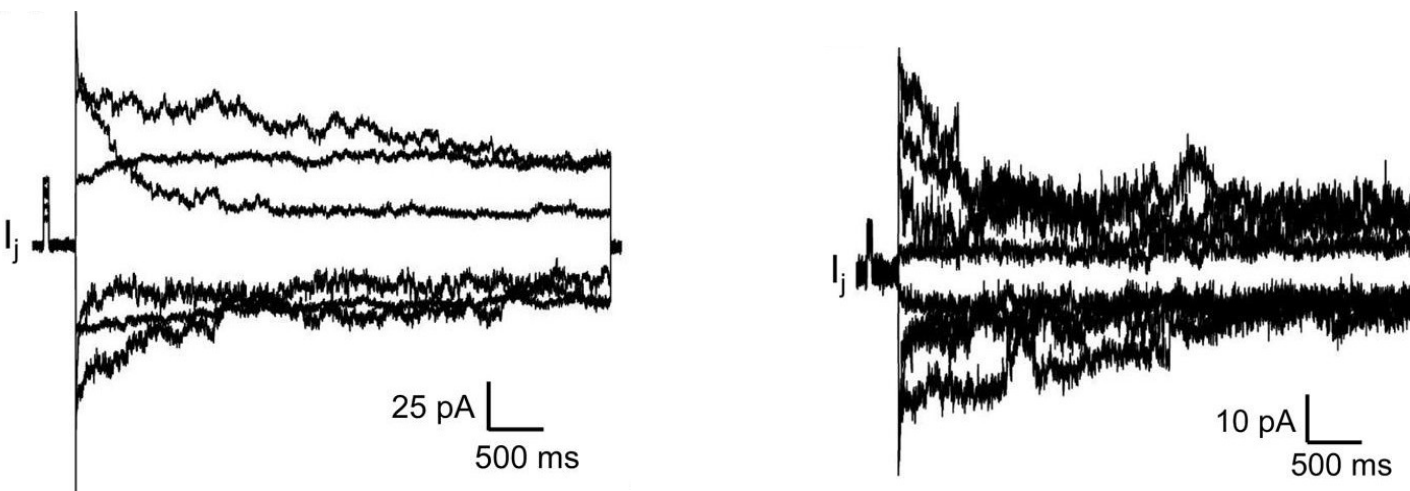
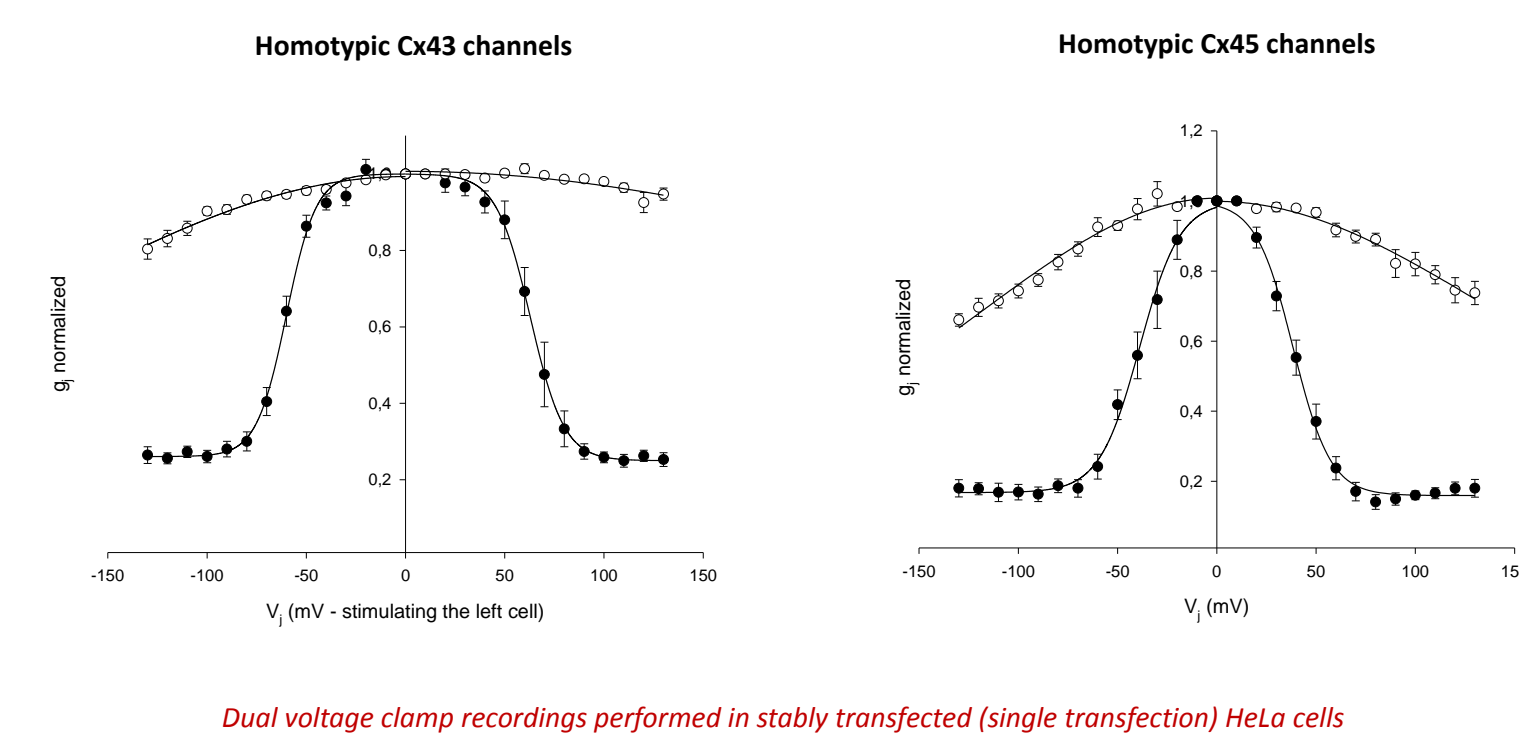


Figure : GJ currents in time, fixed  $V_j$ . Left: Homotypic Cx43/Cx43 cell pairs. Right: Cx43KO/Cx43KO cell pairs.

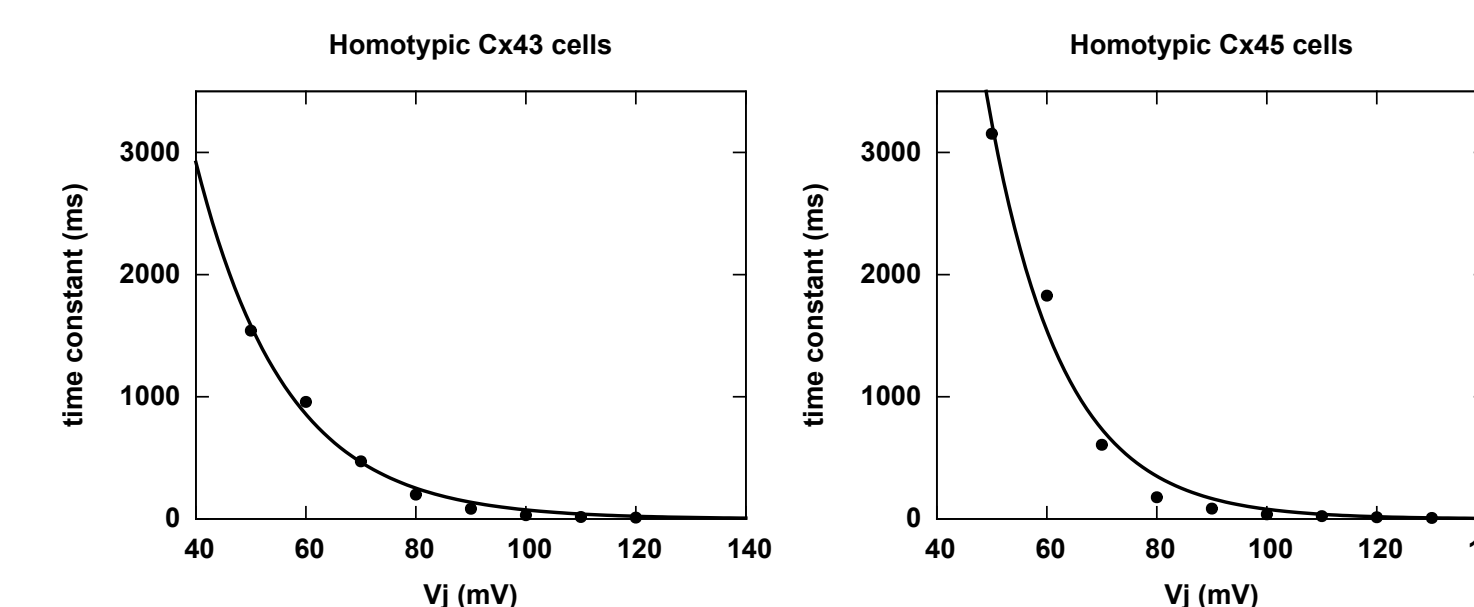
- Conductance:  $G_j(t, V_j) = g_{j,0}g_j(t, V_j)$ .
- The amplitude of the junctional conductance:  $g_{j,0} = 68nS$  for Cx43 cells, i.e.  $g_{j,0} = 2nS$  for Cx43KO.
- Gating variable:  $g_j = g_j(t, V_j)$

$$\frac{dg_j}{dt} = \frac{g_\infty(V_j) - g_j}{\tau_\infty(V_j)}$$

- Fit experimental data to find normalised  $g_\infty(V_j)$ .



- Fit experimental data to find  $\tau_\infty(V_j)$ .



## Macroscopic effects

- Primary cultures of Cx43 WT and Cx43KO ventricular myocytes.
- Macroscopic velocity was calculated from the difference in mean activation time within regions of interest.

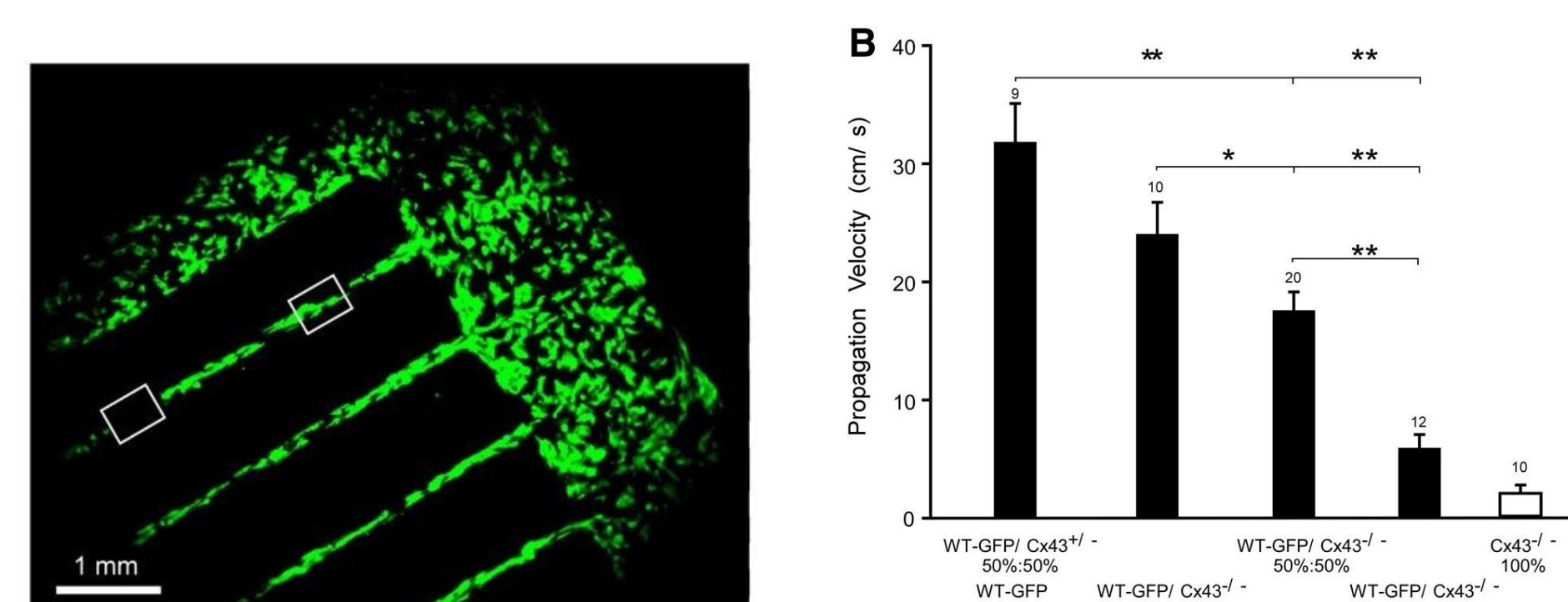


Figure : Left: A mixed patterned cell culture at low magnification. Right: Decrease in velocity of propagation w.r.t. presence of Cx43 cells.

## Assumptions

- Use only homotypic Cx43 and Cx45 GJ channels.
- GJ channels are at the perimeters of the cells, on the membrane.
- The rest of the membrane has only ionic channels. Use Beeler Reuter ionic model.

## 2D/3D mathematical microscopic model

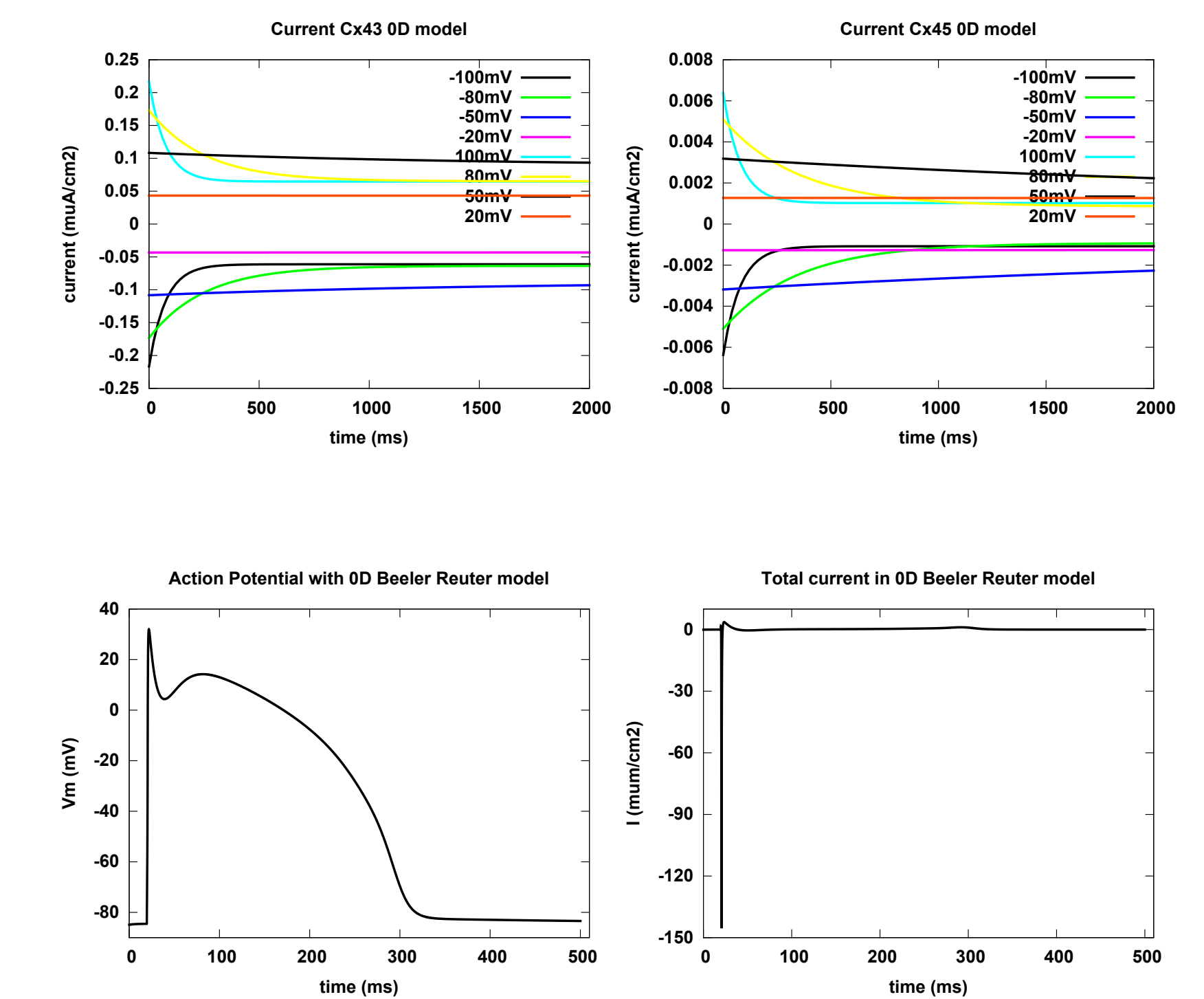
- $\sigma_i, \sigma_e$  intra and extracellular conductivities
- $h$  gating variables for ionic model
- $V_m = u_i - u_e$  transmembrane potential,  $V_j = [u_i]$  transjunctional potential.

$$\begin{aligned} \sigma^i \Delta u^i &= 0, & \text{in } \Omega_i, \\ \sigma^e \Delta u^e &= 0, & \text{in } \Omega_e, \end{aligned}$$

$$\sigma^e \nabla u^e \cdot n = I_{app}(t), \quad \text{on } \partial\Omega_e$$

$$\left. \begin{aligned} \partial_t V_m + I_{ion}(V_m, h) &= \sigma^i \nabla u^i \cdot n, \\ \partial_t V_m + I_{ion}(V_m, h) &= -\sigma^e \nabla u^e \cdot n, \\ \partial_t h &= f_{ion}(V_j, h), \end{aligned} \right\} \text{on } \Gamma_{ion}.$$

$$\left. \begin{aligned} G_j(V_j, g_j) \cdot V_j &= \sigma^i \nabla u^i \cdot n, \\ \partial_t g_j &= f_j(V_m, g_j), \end{aligned} \right\} \text{on } \Gamma_j.$$



## Numerical analysis - 2D

- Tests on:  $2 \times 1$ ,  $4 \times 1$  and  $10 \times 1$  cells, cell size  $100 \times 20 \mu m$ . Mesh step:  $dx = 0.005mm$ .
- Time:  $T = 300ms$ , time step:  $dt = 0.001ms$ . Explicit FE time scheme. Iterative method.
- Execution time on  $10 \times 1$  domain  $\approx 10h$ .

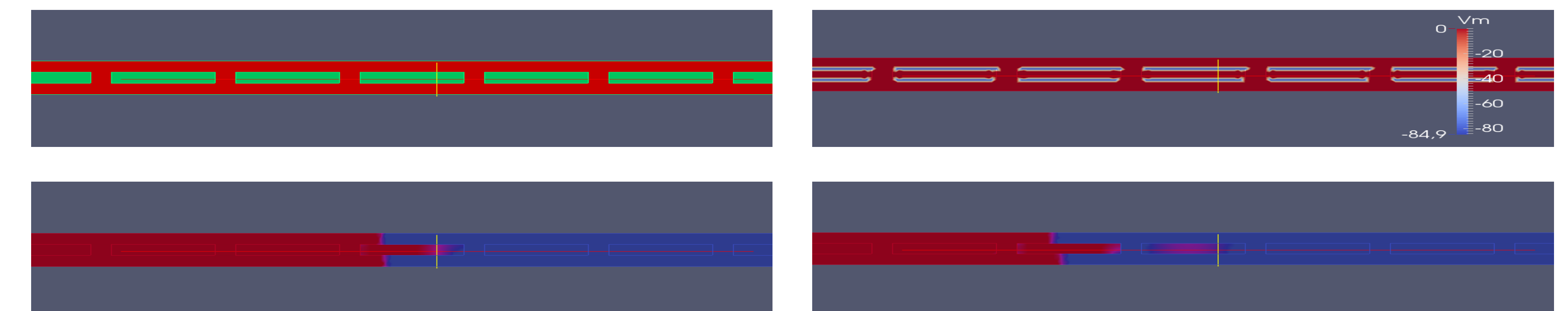


Figure : Up: domain of simulation and initial transmembrane potential. Bottom: propagation of the potential.

## Observations and Future work

- GJ facilitated the propagation in intracellular space.
- Almost NO change in GJ gates - mostly open. Hard to observe dynamics.
- Very small difference due to different GJ channels.

We need:

- Improve the model?
- Longer simulations. Larger domain.
- Try to observe dynamics of GJ channels by applying multiple stimuli.
- Move to HOMOGENISED model!